

AMENDMENT OF THE CLAIMS:

1. (Previously presented) A method for purifying and/or isolating filamentous bacteriophages bacteriophage M13 contained in a solution or a suspension with the capacity for metal chelate formation, the method comprising the steps of:

(a) applying a solution or suspension containing filamentous-bacteriophages bacteriophage M13 onto a metal ions-containing membrane with imidodiacetic acid (IDA) charged with Cu⁺² ions; and

(b) separating the filamentous-bacteriophages bacteriophage M13 from the solution or suspension by affinity chromatography by binding them to the metal ions containing membrane;

—wherein the filamentous-bacteriophages have a molecular weight greater than 1×10^6 daltons (Da).

2. (Cancelled)

3. (Cancelled)

4. (Cancelled)

5. (Cancelled)

6. (Previously presented) The method according to Claim 1, wherein the membrane is a matrix material selected from the group consisting of agaroses, modified agaroses, modified dextrans, polystyrenes, polyethers, polyacrylamides, polyamides, cellulose, modified celluloses, such as cross-linked celluloses, nitrocelluloses, cellulose acetates, silicates and poly(meth)acrylates, polytetrafluoroethylene, polyesters, polyvinyl chlorides, polyvinylidene fluoride, polypropylene, polysulfones and polyethersulfones.

7. (Cancelled)

8. (Cancelled)

9. (Currently amended) The method according to one of Claim 1, wherein ~~a mixture~~ ~~the solution or suspension~~ containing the ~~filamentous~~ ~~bacteriophages~~ ~~bacteriophage M13~~ is subjected to ion exchange chromatography to remove impurities prior to step (a).

10. (Previously presented) The method according to Claim 9, wherein the ion exchange chromatography is performed using an ion exchanger membrane.

11. (Previously presented) The method according to Claim 10, wherein the ion exchanger membrane comprises a matrix material selected from the group consisting of agaroses, modified agaroses, modified dextrans, polystyrenes, polyethers, polyacrylamides, polyamides, cellulose, modified celluloses, such as cross-linked celluloses, nitrocelluloses, cellulose acetates, silicates and poly(meth)acrylates, polytetrafluoroethylenes, polyesters, polyvinyl chlorides, polyvinylidene fluoride, polypropylenes, polysulfones and polyethersulfones.

12. (Previously presented) The method according to Claim 10, wherein the ion exchanger membrane has a pore size in the range of 0.01 to 12 μm , preferably in the range of 0.45 to 7 μm , and especially preferably in the range of 3 to 5 μm .

13. (Currently amended) The method according to Claim 10, wherein the functional groups of the ion exchanger membrane are selected from the group consisting of diethyl aminoethyl (DEAE), ~~2,2'-iminodioethanol~~ (DEA), carboxymethyl (CM), ~~N,N-diethyl-N-(2-hydroxy-1-propyl)ammonioethyl~~ (QA), trimethylamine (TMA), ~~sulfonylmethyl methyl sulfonate~~ (S), sulfopropyl (SP) and phosphate groups.

14. (Previously presented) The method according to Claim 9, wherein the impurities comprise bacterial endotoxins, culture medium components and impurities of culture medium components.

15. (Currently amended) The method according to Claim 1, wherein, prior to step (a) and/or prior to the ion exchange chromatography according to Claim 9, a mixture containing the filamentous bacteriophages bacteriophage M13 is subjected to filtration using a filtration membrane for the removal of additional impurities.

16. (Cancelled)

17. (New) The method according to Claim 1, wherein the membrane has a pore size in the range of 0.01 to 12 μ m.

18. (New) The method according to Claim 1, wherein the membrane has a pore size in the range of 0.45 to 7 μ m.

19. (New) The method according to Claim 1, wherein the membrane has a pore size in the range of 3 to 5 μ m.